

Advance in Research of microRNA in *Caenorhabditis elegans*

Jing Liu, Baofeng Yang,* and Jing Ai*

Department of Pharmacology (the State-Province Key Laboratories of Biomedicine-Pharmaceutics of China, Key Laboratory of Cardiovascular Research, Ministry of Education) Harbin Medical University, No.157 Baojian Road, Nangang District, Harbin, 150081, China

ABSTRACT

microRNA (miRNA) is a family of small, non-coding RNA first discovered as an important regulator of development in *Caenorhabditis elegans* (*C. elegans*). Numerous miRNAs have been found in *C. elegans*, and some of them are well conserved in many organisms. Though, the biologic function of miRNAs in *C. elegans* was largely unknown, more and more studies support the idea that miRNA is an important molecular for *C. elegans*. In this review, we revisit the research progress of miRNAs in *C. elegans* related with development, aging, cancer, and neurodegenerative diseases and compared the function of miRNAs between *C. elegans* and human. *J. Cell. Biochem.* 114: 994–1000, 2013. © 2012 Wiley Periodicals, Inc.

KEY WORDS: microRNAs; *Caenorhabditis elegans*; DEVELOPMENT; LIFESPAN; HUMAN DISEASE

MicroRNA (miRNA) is a family of 21–25 nucleotides small RNA which regulates expression of characterized targets at the post-transcriptional level. The biogenesis of miRNA was well delineated by some reviews [Ambros, 2004; Bartel, 2004; Carthew and Sontheimer, 2009]. In detail, miRNA is initially processed from introns of protein-coding genes or RNA polymerase II (RNAPII) specific transcripts of independent genes. The nascent miRNA transcripts (pri-miRNA) are first cleaved to be precursors (pre-miRNA) by a protein complex Drosha and the double-stranded RNA-binding protein Pasha in the nucleus [Carthew and Sontheimer, 2009; Kim et al., 2009]. Thereafter, the pre-miRNAs are exported to the cytoplasm by exportin-5, and transformed to 21–25 nucleotides mature miRNAs by nuclease Dicer. Finally, mature miRNAs incorporated into miRNA-induced silencing complex (miRISC), and imperfectly bind with complementary sequences in the 3'-untranslated regions (UTRs) of target mRNAs and negatively regulate gene expression through translational inhibition (Fig. 1) [Cullen, 2004; Kim, 2005].

The existence of miRNA in *C. elegans* was first identified by Lee et al. [1993], and the other 154 miRNAs were found in *C. elegans* subsequently by scientists [Grad et al., 2003; Lim et al., 2003]. Noteworthy, recent studies suggested that miRNAs in *C. elegans*

have the similar properties with mammalian. According to the miRBase (release 10.1) database, approximately 62% or 55–62% miRNAs of *C. elegans* relate to *Drosophila* and human [Ibanez-Ventoso et al., 2008], more importantly 34 miRNAs in *C. elegans* are conserved through other species [Lim et al., 2003]. In *C. elegans* the expression levels of miRNAs are varied in different developmental periods [Karp et al., 2011]. The wide conservatism and timing expression suggests miRNAs may play an important role in evolution and development. The present review will highlight advantages of *C. elegans* as a model organism for miRNAs studies and recent findings of miRNAs in *C. elegans*.

THE ADVANTAGES OF *C. elegans* AS A MODEL ORGANISM

The discovery of miRNAs in *C. elegans* has its inevitability. *C. elegans*, as a subtype of nematodes, is the first animal with known whole genome sequence. Its full genome encodes about 20,000 genes, of which at least 40% has homolog genes in the human genome [Sternberg, 2001]. It has two sexes, the hermaphrodite and the male. Based on the sexual character, *C. elegans* display

Grant sponsor: Natural Science Foundation of China; Grant numbers: 81271207, 81070882; Grant sponsor: The Funds for Creative Research Groups of the National Natural Science Foundation of China; Grant number: 81121003; Grant sponsor: Outstanding Youth Foundation of Heilongjiang Province; Grant number: JC200904.

*Correspondence to: Jing Ai or Baofeng Yang, Department of Pharmacology, Harbin Medical University, No.157 Baojian Road, Nangang District, Harbin 150081, China. E-mail: a.z.hrbmu@gmail.com; yangbf@ems.hrbmu.edu.cn

Manuscript Received: 28 October 2012; Manuscript Accepted: 1 November 2012

Accepted manuscript online in Wiley Online Library (wileyonlinelibrary.com): 13 November 2012

DOI 10.1002/jcb.24448 • © 2012 Wiley Periodicals, Inc.

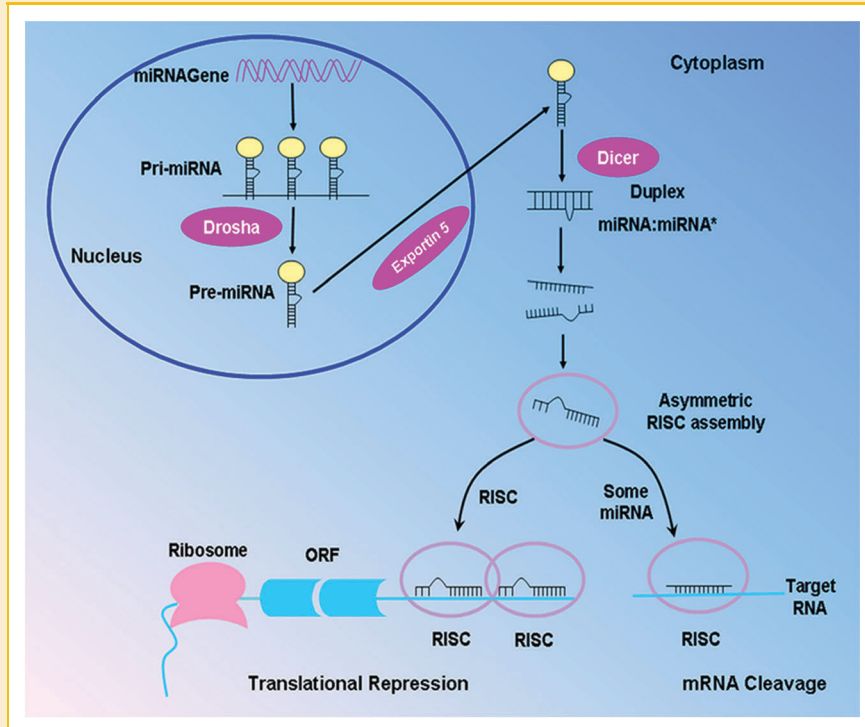


Fig. 1. Model of the biogenesis and post-transcriptional suppression of miRNAs. The pri-miRNA transcripts are first processed into pre-miRNA by Drosha inside the nucleus. Pre-miRNAs are transported into the cytoplasm by Exportin 5 and then become miRNA:miRNA* duplexes by Dicer. Next, one strand of the miRNA:miRNA* duplex is assembled into the miRNA-induced silencing complex (miRISC). Finally, miRNA acts on its target by translational repression or mRNA cleavage.

unparalleled advantages in genetic studies. Hermaphrodite *C. elegans* are common used to conduct genetic mating and analysis due to their breed true by self-fertilization and keep the traits without mating [Strange, 2006]. *C. elegans* also used to study developmental mechanism because of their clear developmental cycle (embryogenesis and four larval stages: L1-L4), which was punctuated by molts [Luo, 2004]. In a favorable environment, larval development is rapid and continuous [Sulston and Horvitz, 1977]. However, in unfavorable environment *C. elegans* will enter a developmental arrested, long-lived, and highly stress-resistant stage called dauer diapause [Cassada and Russell, 1975]. Furthermore, recent studies have found that some miRNA and proteins in dauer larvae were different from non-dauer animals in various developmental stages [Jones et al., 2001; Wang and Kim, 2003]. These features of *C. elegans* provided an excellent animal model to study mechanisms of cell differentiation, development, and aging. It is worth mentioning that there are many landmark discoveries using the small worm. In 1974, Brenner used Ethyl Methanesulphonate (EMS) chemical mutagenesis to induce wild-type *C. elegans* and established ~300 mutants with characterized behavior and morphology [Brenner, 1974]. It laid the foundation for *C. elegans* in the animal behavior and development genetic study. In 1977, Sulston used the differential interfering contrast (DIC) technology to study different phenotypes of cell characteristics in *C. elegans* and painted a unique cell fates lineage diagram [Sulston and Horvitz, 1977; Sulston et al., 1983], which provided a platform for scientists to study the genetic development regulatory mechanism on the

single cell in *C. elegans*. Based on above work, Horvitz cloned dozen genes that regulated the programmed cell death in *C. elegans* and the following studies found these genes were similar with mammals [Ellis and Horvitz, 1986; Horvitz, 1999]. In 2002, Sydney Brenner shared the Nobel Prize in Physiology and Medicine with John Sulston and H. Robert Horvitz for their discovery of genes in *C. elegans* that regulate organ development and programmed cell death. These achievements display the irreplaceable position of *C. elegans* in the field of biomedical research.

FUNCTION OF miRNAs In *C. elegans* DEVELOPMENT

Animal development is a complex, strictly regulated process. Recent studies discovered miRNAs important roles in development. It was report that 12 miRNAs expression increases dramatically and eight miRNAs expression decreases dramatically in the lifespan from L1-L4 [Karp et al., 2011]. Among these miRNAs, the lin-4 and let-7 family are well studied. The lin-4 was found to control the L1 to L2 development of *C. elegans* by targeting on lin-14 and lin-28 mRNAs and then the two mRNAs regulate hbl-1 expression directly [Ambros, 1989; Wightman et al., 1993]. The let-7 was reported to control cell fate of hypodermal in the time of late-larval development [Reinhart et al., 2000]. Over-expression of let-7 can prevent the development of *C. elegans* from the L4 to adult by directly inhibiting the expression of lin-41, hbl-1, daf-12, and pha-4

mRNAs. These mRNAs were further found to regulate the development of *C. elegans* by depressing the transcription factor lin-29 [Slack et al., 2000; Lin et al., 2003; Grosshans et al., 2005]. Furthermore, miR-48, miR-84, and miR-241 were found to participate in the L2 to L3 development of *C. elegans* by targeting on hbl-1 mRNA. In addition, miR-48 and miR-84 also acted on cessation of the larval molting cycle at the adult stage, but their target genes are unknown (Fig. 2) [Abbott et al., 2005].

FUNCTION OF miRNAs IN *C. elegans* LIFESPAN

At the achievement of the larval development, aging is following and along with miRNAs change. de Lencastre et al. [2010] reported that comparing with Day 0 of adulthood wild-type *C. elegans*, 7 miRNAs were significantly up-regulated, and 23 miRNAs greatly down-regulated on day 10, suggesting miRNAs may involved in aging pathways to regulate the lifespan of *C. elegans*.

Insulin/IGF-1 signaling (IIS) is the first pathway identified in regulating *C. elegans* lifespan [Kenyon et al., 1993]. In this pathway, DAF-2 (insulin receptor-like protein) is the key component to control the lifespan of *C. elegans* by regulating the expression of both transcription factor abnormal dauer formation-16 (DAF-16) and heat shock factor-1 (HSF-1) [Lin et al., 1997; Hsu et al., 2003]. The loss of daf-2 function in *C. elegans* is considered to increase lifespan, while gain of daf-16 function is required for the longevity

which can be antagonized by daf-2 in wild-type *C. elegans* [Lin et al., 2001]. In 2005, Boehm and Slack first reported that over-expression of lin-4 can induce a longevity in *C. elegans*, whereas loss of lin-4 function has an opposite phenomenon. Thereafter, they found the lin-4 induced longevity by targeting on the lin-14 which act on the daf-16 and hsf-1 of IIS pathway [Boehm and Slack, 2005; Boehm and Slack, 2006]. Lin-4 is the first and a classical example of miRNA to regulate lifespan. These phenomena suggest miRNAs could regulate *C. elegans* lifespan by affecting different stage. However, whether miRNAs affect the lifespan of *C. elegans* by direct acting on Insulin/IGF-1 signaling pathway is unknown.

Dietary Restriction (DR) is another molecular mechanism in *C. elegans* lifespan regulation [Lakowski and Hekimi, 1998]. Studies indicated that the extension of lifespan in the eat-2 (a nicotinic acetylcholine receptor subunit that acts in the pharyngeal muscle) mutant *C. elegans* by DR was dependent on a transcription factor named by pharynx development defect-4 (PHA-4) [Panowski et al., 2007] rather than DAF-16 [Houthoofd et al., 2003]. Interest, based on the miRNAs databases (TargetScan), pha-4 is a potential target of let-7. However, there is no report that let-7 can affect lifespan through down-regulating pha-4 in aging *C. elegans* until now. Therefore, whether miRNAs involved in DR pathway is still unclear up-to-now.

It is well known that oxidative stress can affect lifespan [Kenyon, 2005]. However, whether aging-associated miRNAs could influence *C. elegans* lifespan through affecting the response of *C. elegans* to

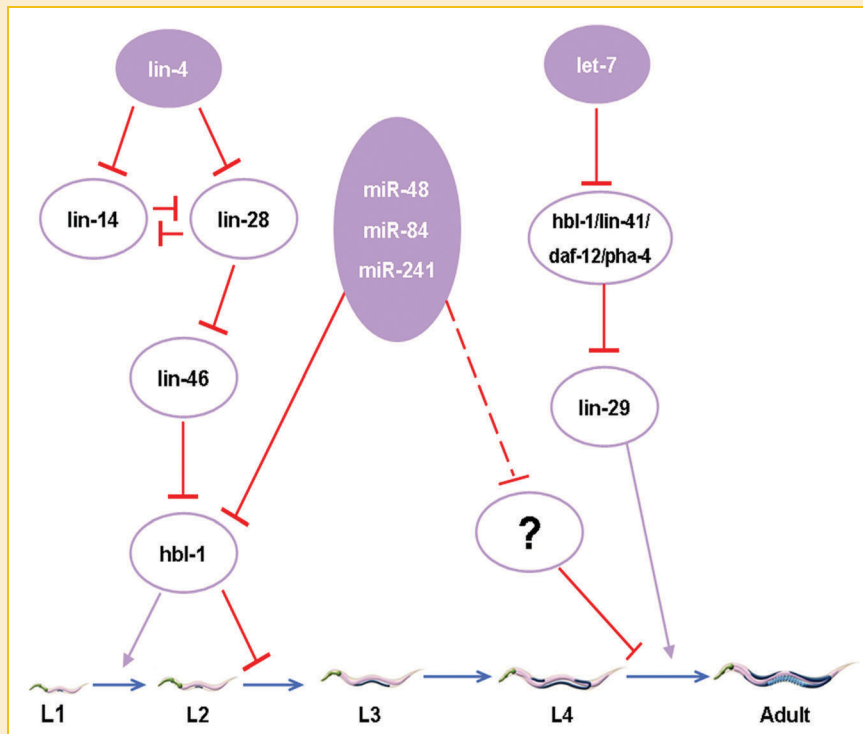


Fig. 2. The function of the miRNAs in *C. elegans* development. The lin-4 miRNA targets lin-14 and lin-28, which interact in early larval development. Lin-46 acts downstream of lin-28 and regulates the expression of hbl-1. The let-7 miRNA targets the lin-41, hbl-1, daf-12, and pha-4 mRNAs, which lead to the expression of the transcription factor lin-29. miR-48, miR-84, and miR-241 which are let-7-like miRNAs regulate the hbl-1 mRNA at the early development time. The dotted line represents a possible role of mir-48, mir-84, and mir-241 during development.

stress is unknown. Previous studies found that deletion of miR-239 enhanced resistance to both heat stress and oxidative stress finally resulted in longevity, while deletion of miR-71 was converse. In contrast, deletions of miR-238 and miR-246 induce increased sensitivity to oxidation and heat stress, respectively [de Lencastre et al., 2010]. However, the exact mechanism is still unknown. The transcriptional factor *skn-1* plays a key role in the oxidative stress pathway, and is also necessary for intestinal development in *C. elegans* [An and Blackwell, 2003; Park et al., 2009]. Though *skn-1* is a target for all miR-71, miR-238, and miR-239, whether these miRNAs act on *skn-1* and regulate the *C. elegans* response to stress and lifespan need to be further studied (Fig. 3).

miRNA AND DISEASE: *C. elegans* TO HUMAN

Since human share the common laws of development and differentiation with *C. elegans*, studies about miRNAs in *C. elegans* would provide invaluable hints to understand the role of miRNAs on physiological and pathophysiological process in human.

In human, inappropriate cell proliferation and differentiation are reminiscent of cancer, and recent studies indicated that many miRNAs were involved in development of various human malignancies [Lu et al., 2005]. Importantly, some miRNAs, which are associated with organism development, cell proliferation, and

differentiation in *C. elegans*, are found to be potential predictors for tumors in human. For example, *let-7* is found associated with cancer. Vitro experiments have revealed that *let-7* is reduced frequently in lung cancer and over-expression of *let-7* can inhibit the growth of lung cancer cells in human [Takamizawa et al., 2004]. Whereas, though miR-34 up-regulated in both *C. elegans* and mammalian cells in post-radiation, studies found that the up-regulated miR-34 level was p53-independent in *C. elegans* [Kato et al., 2009], but p53-dependent in human [Hermeking, 2009]. The difference suggests that the role of miRNAs in *C. elegans* could provide valuable information for human diseases but not completely used for human diseases. Besides *let-7* and miR-34, other miRNAs involved in *C. elegans* development are also associated with cancer (Table I).

C. elegans is also an excellent animal used for studying neurodegenerative disease, such as Alzheimer's disease (AD) and Parkinson's disease (PD). In AD patient, dysfunction of the amyloid precursor protein (APP) is one of the major risk factor [Selkoe, 2007]. In *C. elegans*, *apl-1* is an APP-related gene, which is conserved in evolution [Daigle and Li, 1993]. Though, no study reported the directly regulation action of miRNAs on *apl-1*, *apl-1* was found have significant genetic interactions with *let-7* family [Niwa et al., 2008]. These results indicate that *apl-1* expression is temporally regulated by miRNAs. It may provide new insights into the time dependent progression of AD. In PD-associated *C. elegans* model, miR-64 and miR-65 are co-expressed in α -synuclein transgenic strain and *cat-1*

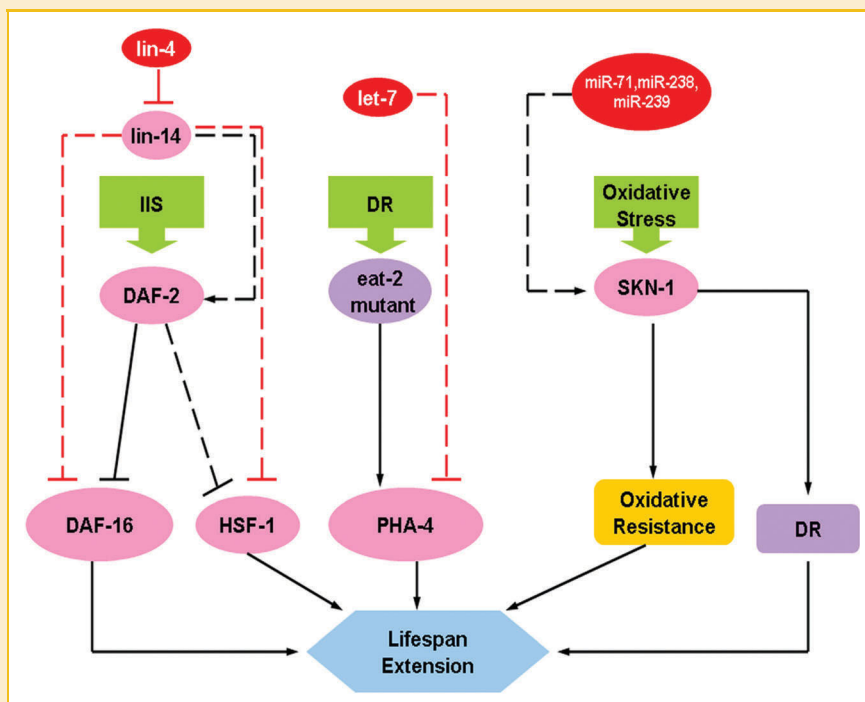


Fig. 3. Pathways that influence lifespan extension in response to miRNAs regulation. In IIS pathway, DAF-16 is antagonized by the DAF-2. The low level of lin-14 expression enhances the longevity promoters DAF-16 and HSF-1 expression. In DR pathway, defection of *eat-2* enhances expression of transcription factor PHA-4 which is required to mediate DR induced longevity specifically and it is also the validated target of the *let-7*. SKN-1 is a transcription factor required for responding to oxidative stress in intestine, and also required for DR-induced lifespan extension [Bishop and Guarente, 2007]. The miR-71, miR-238, and miR-239 are involved in stress resistance suggesting possibility of these miRNAs function through stress resistance to mediate lifespan in *C. elegans*. Dotted lines represent possible roles in these pathways.

TABLE I. Some miRNAs Function in *C. elegans* Development Associate With Cancer

miRNA	Target(s)	Function	Homology	Property	Malignancy	Reference
let-7, miR-48, miR-84, miR-241	lin4, hbl-1, daf-12, pha-4, let-60	Stem cell difference, cessation of molting	Hs-let-7, Hs-miR-98, Hs-miR-196	TS	CLL, lymphoma, gastric, lung, prostate, breast, ovarian, colon, leiomyoma, melanoma	[Reinhart et al., 2000; Slack et al., 2000; Abrahante et al., 2003; Lin et al., 2003; Abbott et al., 2005; Alvarez-Garcia and Miska, 2005; Grosshans et al., 2005; Li et al., 2005; Calin et al., 2008; Roush and Slack, 2008; Peter, 2009; Garofalo and Croce, 2010; O'Day and Lal, 2010; Osada and Takahashi, 2010]
miR-34a, miR-34b, miR-34c	SIRT1	Cell death	Hs-miR-34	TS	CLL, lymphoma, pancreatic, colon, lung, neuroblastoma, glioblastoma	[Alvarez-Garcia and Miska, 2005; Calin et al., 2008; Cole et al., 2008; Garofalo and Croce, 2010; Li et al., 2009; Wiggins et al., 2010; Yamakuchi et al., 2008; Yamakuchi and Lowenstein, 2009]
lin-4, miR-273	lin-14, lin-28, die-1	Stem cell difference, left-right asymmetry	Hs-miR-125b	TS	breast, ovarian, lung, prostate, urothelium, colorectum	[Lee et al., 1993; Wightman et al., 1993; Negrini et al., 1995; Rasio et al., 1995; Chang et al., 2003; Alvarez-Garcia and Miska, 2005; Iorio et al., 2005; Ozen et al., 2008; Veerla et al., 2009; Nishida et al., 2011]
lys-6	cog-1	Left-right asymmetry				[Johnston and Hobert, 2003; Alvarez-Garcia and Miska, 2005]

miRNAs that are down-regulated in malignancies are named as tumor-suppressor (TS), but their function in malignancy is not all experimentally validated.

strain. Additionally members of let-7 family are co-expressed in the α -synuclein transgenic strain and pdr-1 strain [Asikainen et al., 2010]. These results suggest that different miRNAs may express on different PD models of *C. elegans* and lead to different PD pathogenesis.

FUTURE DIRECTIONS

Up-to-today, there are some of miRNAs that could regulate organism development, cell proliferation, and differentiation as well as lifespan in *C. elegans*, but the function of miRNAs in *C. elegans* is largely unknown. Therefore, there will be an extensive future in miRNAs research in *C. elegans*. Since the homology of miRNAs in *C. elegans* and mammals, it will be a huge challenge to study the correlation of miRNAs between *C. elegans* and mammals even human. We believe *C. elegans* will help us further understand miRNAs function and may discover new areas of small RNA world.

REFERENCES

Abbott AL, Alvarez-Saavedra E, Miska EA, Lau NC, Bartel DP, Horvitz HR, Ambros V. 2005. The let-7 MicroRNA family members mir-48, mir-84, and mir-241 function together to regulate developmental timing in *Caenorhabditis elegans*. *Dev Cell* 9:403–414.

Abrahante JE, Daul AL, Li M, Volk ML, Tennessen JM, Miller EA, Rougvie AE. 2003. The *Caenorhabditis elegans* hunchback-like gene lin-57/hbl-1 controls developmental time and is regulated by microRNAs. *Dev Cell* 4:625–637.

Alvarez-Garcia I, Miska EA. 2005. MicroRNA functions in animal development and human disease. *Development* 132:4653–4662.

Ambros V. 1989. A hierarchy of regulatory genes controls a larva-to-adult developmental switch in *C. elegans*. *Cell* 57:49–57.

Ambros V. 2004. The functions of animal microRNAs. *Nature* 431:350–355.

An JH, Blackwell TK. 2003. SKN-1 links *C. elegans* mesodermal specification to a conserved oxidative stress response. *Genes Dev* 17:1882–1893.

Asikainen S, Rudgalvyte M, Heikkinen L, Louhiranta K, Lakso M, Wong G, Nass R. 2010. Global microRNA expression profiling of *Caenorhabditis elegans* Parkinson's disease models. *J Mol Neurosci* 41:210–218.

Bartel DP. 2004. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* 116:281–297.

Bishop NA, Guarente L. 2007. Two neurons mediate diet-restriction-induced longevity in *C. elegans*. *Nature* 447:545–549.

Boehm M, Slack F. 2005. A developmental timing microRNA and its target regulate life span in *C. elegans*. *Science* 310:1954–1957.

Boehm M, Slack FJ. 2006. MicroRNA control of lifespan and metabolism. *Cell Cycle* 5:837–840.

Brenner S. 1974. The genetics of *Caenorhabditis elegans*. *Genetics* 77:71–94.

Calin GA, Cimmino A, Fabbri M, Ferracin M, Wojcik SE, Shimizu M, Taccioli C, Zanesi N, Garzon R, Aqeilan RI, Alder H, Volinia S, Rassenti L, Liu X, Liu CG, Kipps TJ, Negrini M, Croce CM. 2008. MiR-15a and miR-16-1 cluster functions in human leukemia. *Proc Natl Acad Sci USA* 105:5166–5171.

Carthew RW, Sontheimer EJ. 2009. Origins and mechanisms of miRNAs and siRNAs. *Cell* 136:642–655.

Cassada RC, Russell RL. 1975. The dauerlarva, a post-embryonic developmental variant of the nematode *Caenorhabditis elegans*. *Dev Biol* 46:326–342.

- Chang S, Johnston RJ, Jr., Hobert O. 2003. A transcriptional regulatory cascade that controls left/right asymmetry in chemosensory neurons of *C. elegans*. *Genes Dev* 17:2123–2137.
- Cole KA, Attiyeh EF, Mosse YP, Laquaglia MJ, Diskin SJ, Brodeur GM, Maris JM. 2008. A functional screen identifies miR-34a as a candidate neuroblastoma tumor suppressor gene. *Mol Cancer Res* 6:735–742.
- Cullen BR. 2004. Transcription and processing of human microRNA precursors. *Mol Cell* 16:861–865.
- Daigle I, Li C. 1993. *apl-1*, a *Caenorhabditis elegans* gene encoding a protein related to the human beta-amyloid protein precursor. *Proc Natl Acad Sci USA* 90:12045–12049.
- de Lencastre A, Pincus Z, Zhou K, Kato M, Lee SS, Slack FJ. 2010. MicroRNAs both promote and antagonize longevity in *C. elegans*. *Curr Biol* 20:2159–2168.
- Ellis HM, Horvitz HR. 1986. Genetic control of programmed cell death in the nematode *C. elegans*. *Cell* 44:817–829.
- Garofalo M, Croce CM. 2010. microRNAs: Master regulators as potential therapeutics in cancer. *Annu Rev Pharmacol Toxicol* 51:25–43.
- Grad Y, Aach J, Hayes GD, Reinhart BJ, Church GM, Ruvkun G, Kim J. 2003. Computational and experimental identification of *C. elegans* microRNAs. *Mol Cell* 11:1253–1263.
- Grosshans H, Johnson T, Reinert KL, Gerstein M, Slack FJ. 2005. The temporal patterning microRNA *let-7* regulates several transcription factors at the larval to adult transition in *C. elegans*. *Dev Cell* 8:321–330.
- Hermeking H. 2009. The miR-34 family in cancer and apoptosis. *Cell Death Differ* 17:193–199.
- Horvitz HR. 1999. Genetic control of programmed cell death in the nematode *Caenorhabditis elegans*. *Cancer Res* 59:1701s–1706s.
- Houthoofd K, Braeckman BP, Johnson TE, Vanfleteren JR. 2003. Life extension via dietary restriction is independent of the Ins/IGF-1 signalling pathway in *Caenorhabditis elegans*. *Exp Gerontol* 38:947–954.
- Hsu AL, Murphy CT, Kenyon C. 2003. Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science* 300:1142–1145.
- Ibanez-Ventoso C, Vora M, Driscoll M. 2008. Sequence relationships among *C. elegans*, *D. melanogaster* and human microRNAs highlight the extensive conservation of microRNAs in biology. *PLoS One* 3:e2818.
- Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, Magri E, Pedriali M, Fabbri M, Campiglio M, Menard S, Palazzo JP, Rosenberg A, Musiani P, Volinia S, Nenci I, Calin GA, Querzoli P, Negrini M, Croce CM. 2005. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res* 65:7065–7070.
- Johnston RJ, Hobert O. 2003. A microRNA controlling left/right neuronal asymmetry in *Caenorhabditis elegans*. *Nature* 426:845–849.
- Jones SJ, Riddle DL, Pouzyrev AT, Velculescu VE, Hillier L, Eddy SR, Stricklin SL, Baillie DL, Waterston R, Marra MA. 2001. Changes in gene expression associated with developmental arrest and longevity in *Caenorhabditis elegans*. *Genome Res* 11:1346–1352.
- Karp X, Hammell M, Ow MC, Ambros V. 2011. Effect of life history on microRNA expression during *C. elegans* development. *RNA* 17:639–651.
- Kato M, Paranjape T, Muller RU, Nallur S, Gillespie E, Keane K, Esquela-Kerscher A, Weidhaas JB, Slack FJ. 2009. The miR-34 microRNA is required for the DNA damage response in vivo in *C. elegans* and in vitro in human breast cancer cells. *Oncogene* 28:2419–2424.
- Kenyon C. 2005. The plasticity of aging: Insights from long-lived mutants. *Cell* 120:449–460.
- Kenyon C, Chang J, Gensch E, Rudner A, Tabtiang R. 1993. A *C. elegans* mutant that lives twice as long as wild type. *Nature* 366:461–464.
- Kim VN. 2005. MicroRNA biogenesis: Coordinated cropping and dicing. *Nat Rev Mol Cell Biol* 6:376–385.
- Kim VN, Han J, Siomi MC. 2009. Biogenesis of small RNAs in animals. *Nat Rev Mol Cell Biol* 10:126–139.
- Lakowski B, Hekimi S. 1998. The genetics of caloric restriction in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 95:13091–13096.
- Lee RC, Feinbaum RL, Ambros V. 1993. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 75:843–854.
- Li M, Jones-Rhoades MW, Lau NC, Bartel DP, Rougvie AE. 2005. Regulatory mutations of *mir-48*, a *C. elegans* *let-7* family MicroRNA, cause developmental timing defects. *Dev Cell* 9:415–422.
- Li N, Fu H, Tie Y, Hu Z, Kong W, Wu Y, Zheng X. 2009. miR-34a inhibits migration and invasion by down-regulation of c-Met expression in human hepatocellular carcinoma cells. *Cancer Lett* 275:44–53.
- Lim LP, Lau NC, Weinstein EG, Abdelhakim A, Yekta S, Rhoades MW, Burge CB, Bartel DP. 2003. The microRNAs of *Caenorhabditis elegans*. *Genes Dev* 17:991–1008.
- Lin K, Dorman JB, Rodan A, Kenyon C. 1997. *daf-16*: An HNF-3/forkhead family member that can function to double the life-span of *Caenorhabditis elegans*. *Science* 278:1319–1322.
- Lin K, Hsin H, Libina N, Kenyon C. 2001. Regulation of the *Caenorhabditis elegans* longevity protein DAF-16 by insulin/IGF-1 and germline signaling. *Nat Genet* 28:139–145.
- Lin SY, Johnson SM, Abraham M, Vella MC, Pasquinelli A, Gamberi C, Gottlieb E, Slack FJ. 2003. The *C. elegans* hunchback homolog, *hbl-1*, controls temporal patterning and is a probable microRNA target. *Dev Cell* 4:639–650.
- Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA, Downing JR, Jacks T, Horvitz HR, Golub TR. 2005. MicroRNA expression profiles classify human cancers. *Nature* 435:834–838.
- Luo Y. 2004. Long-lived worms and aging. *Redox Rep* 9:65–69.
- Negrini M, Rasio D, Hampton GM, Sabbioni S, Rattan S, Carter SL, Rosenberg AL, Schwartz GF, Shiloh Y, Cavenee WK, Croce CM. 1995. Definition and refinement of chromosome 11 regions of loss of heterozygosity in breast cancer: Identification of a new region at 11q23.3. *Cancer Res* 55:3003–3007.
- Nishida N, Yokobori T, Mimori K, Sudo T, Tanaka F, Shibata K, Ishii H, Doki Y, Kuwano H, Mori M. 2011. MicroRNA miR-125b is a prognostic marker in human colorectal cancer. *Int J Oncol* 38:1437–1443.
- Niwa R, Zhou F, Li C, Slack FJ. 2008. The expression of the Alzheimer's amyloid precursor protein-like gene is regulated by developmental timing microRNAs and their targets in *Caenorhabditis elegans*. *Dev Biol* 315:418–425.
- O'Day E, Lal A. 2010. MicroRNAs and their target gene networks in breast cancer. *Breast Cancer Res* 12:201.
- Osada H, Takahashi T. 2010. *let-7* and miR-17-92: Small-sized major players in lung cancer development. *Cancer Sci* 102:9–17.
- Ozen M, Creighton CJ, Ozdemir M, Ittmann M. 2008. Widespread deregulation of microRNA expression in human prostate cancer. *Oncogene* 27:1788–1793.
- Panowski SH, Wolff S, Aguilaniu H, Durieux J, Dillin A. 2007. PHA-4/Foxa mediates diet-restriction-induced longevity of *C. elegans*. *Nature* 447:550–555.
- Park SK, Tedesco PM, Johnson TE. 2009. Oxidative stress and longevity in *Caenorhabditis elegans* as mediated by SKN-1. *Aging Cell* 8:258–269.
- Peter ME. 2009. *Let-7* and miR-200 microRNAs: Guardians against pluripotency and cancer progression. *Cell Cycle* 8:843–852.
- Rasio D, Negrini M, Manenti G, Dragani TA, Croce CM. 1995. Loss of heterozygosity at chromosome 11q in lung adenocarcinoma: Identification of three independent regions. *Cancer Res* 55:3988–3991.

- Reinhart BJ, Slack FJ, Basson M, Pasquinelli AE, Bettinger JC, Rougvie AE, Horvitz HR, Ruvkun G. 2000. The 21-nucleotide let-7 RNA regulates developmental timing in *Caenorhabditis elegans*. *Nature* 403:901–906.
- Roush S, Slack FJ. 2008. The let-7 family of microRNAs. *Trends Cell Biol* 18:505–516.
- Selkoe DJ. 2007. Developing preventive therapies for chronic diseases: Lessons learned from Alzheimer's disease. *Nutr Rev* 65:S239–S243.
- Slack FJ, Basson M, Liu Z, Ambros V, Horvitz HR, Ruvkun G. 2000. The lin-41 RBCC gene acts in the *C. elegans* heterochronic pathway between the let-7 regulatory RNA and the LIN-29 transcription factor. *Mol Cell* 5:659–669.
- Sternberg PW. 2001. Working in the post-genomic *C. elegans* world. *Cell* 105:173–176.
- Strange K. 2006. An overview of *C. elegans* biology. *Methods Mol Biol* 351:1–11.
- Sulston JE, Horvitz HR. 1977. Post-embryonic cell lineages of the nematode, *Caenorhabditis elegans*. *Dev Biol* 56:110–156.
- Sulston JE, Schierenberg E, White JG, Thomson JN. 1983. The embryonic cell lineage of the nematode *Caenorhabditis elegans*. *Dev Biol* 100:64–119.
- Takamizawa J, Konishi H, Yanagisawa K, Tomida S, Osada H, Endoh H, Harano T, Yatabe Y, Nagino M, Nimura Y, Mitsudomi T, Takahashi T. 2004. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res* 64:3753–3756.
- Veerla S, Lindgren D, Kvist A, Frigyesi A, Staaf J, Persson H, Liedberg F, Chebil G, Gudjonsson S, Borg A, Mansson W, Rovira C, Hoglund M. 2009. MiRNA expression in urothelial carcinomas: Important roles of miR-10a, miR-222, miR-125b, miR-7 and miR-452 for tumor stage and metastasis, and frequent homozygous losses of miR-31. *Int J Cancer* 124:2236–2242.
- Wang J, Kim SK. 2003. Global analysis of dauer gene expression in *Caenorhabditis elegans*. *Development* 130:1621–1634.
- Wiggins JF, Ruffino L, Kelnar K, Omotola M, Patrawala L, Brown D, Bader AG. 2010. Development of a lung cancer therapeutic based on the tumor suppressor microRNA-34. *Cancer Res* 70:5923–5930.
- Wightman B, Ha I, Ruvkun G. 1993. Posttranscriptional regulation of the heterochronic gene lin-14 by lin-4 mediates temporal pattern formation in *C. elegans*. *Cell* 75:855–862.
- Yamakuchi M, Ferlito M, Lowenstein CJ. 2008. miR-34a repression of SIRT1 regulates apoptosis. *Proc Natl Acad Sci USA* 105:13421–13426.
- Yamakuchi M, Lowenstein CJ. 2009. MiR-34, SIRT1 and p53: The feedback loop. *Cell Cycle* 8:712–715.